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Pattern of carbon dioxide production and retention is similar in adult pigs when fed hourly, but not when fed a single meal

Soenke Moehn¹, Robert FP Bertolo^{1,2}, Paul B Pencharz^{1,3} and Ronald O Ball*¹

Address: ¹Department of Agricultural, Food and Nutritional Science, University of Alberta, Edmonton, AB, T6G 2P5 Canada, ²Department of Biochemistry, Memorial University of Newfoundland, St. John's, NL, A1B 3X9 Canada and ³Research Institute, Hospital for Sick Children, Toronto, ON, Canada, M5G 1X8

Email: Soenke Moehn - smoehn@ualberta.ca; Robert FP Bertolo - rbertolo@mun.ca; Paul B Pencharz - paul.pencharz@sickkids.on.ca; Ronald O Ball* - ron.ball@ualberta.ca

* Corresponding author

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Abstract

Background: The understanding of bicarbonate kinetics and CO₂ retention in the body is necessary to conduct amino acid tracer oxidation studies in both humans and laboratory animals. Significant metabolic activity is associated with eating which can affect bicarbonate steady state kinetics. A study was conducted to assess the impact of feeding regimen on the recovery of labelled bicarbonate and energy expenditure in adult female pigs (sows). Five catheterized sows (235 ± 5 kg) were fed semi-synthetic diets as: a single meal 2 h into the infusion after an overnight fast, or in eight hourly meals starting 2 h before the infusion. Oxygen consumption, CO₂ production and ¹⁴CO₂ recovery (ie fraction not retained) were determined during primed, constant intravenous infusions of NaH¹⁴CO₃.

Results: The ¹⁴CO₂ recovery (%) after fasting (58.1 ± 4.8) was lower than that after single meal feeding (78.8 ± 5.9) or hourly meal feeding (81.0 ± 2.6, *P* = 0.03). CO₂ production correlated with ¹⁴CO₂ recovery during hourly feeding (*r* = 0.40, *P* = 0.01); this relationship was not significant after single meal feeding (*P* = 0.30), probably due to physical activity-associated CO₂ production.

Conclusions: The correlation of CO₂ retention factors with CO₂ production during hourly feeding suggests that this regimen should be preferred for future amino acid kinetics studies.

Background

Traditionally, measures of energy and protein metabolism have rarely been studied simultaneously in the same animal although there is a need to do so. We constructed an experimental protocol that allows the measurement of energy expenditure at the same time as the measurement of amino acid oxidation for studies in amino acid requirements, protein kinetics and energy metabolism in adult pigs.

The study of amino acid requirements using an amino acid oxidation method necessitates the knowledge of the degree of retention of labelled CO₂ generated during the oxidation of the labelled amino acid. Although bicarbonate kinetics and CO₂ retention are well documented for humans [1], parenterally-fed piglets [2] and chickens [3], no data were found for adult sows, which we intend to use to answer questions in both human and pig nutrition. Although the retention of CO₂ is dependent on the

physiological state (i.e. post-absorptive or postprandial) and on the type and extent of exercise in humans [1], comparable observations in pigs are lacking. The retention of CO₂ may be linked to the total production of CO₂ by the animal, which has been shown to fluctuate with physical activity and digestion [4]. Because an influence of feeding and activity on the retention of CO₂ and on energy metabolism is expected, it is necessary to identify an appropriate experimental protocol to avoid biased measurements of amino acid oxidation and energy expenditure and to maintain the comparability of measurements made between animals.

The objectives of this experiment were: 1) to determine the CO₂ retention in adult sows during the post-absorptive state and the postprandial state when fed either a single meal or hourly meals, 2) to assess the effect of these feeding regimens on CO₂ and heat production, and 3) to describe the relationship between CO₂ retention and CO₂ production.

Results

For the calibration of the respiration systems, the mean recoveries of ¹⁴CO₂ and CO₂ were 101.8% (SD 3.1, *n* = 16) and 105.2% (SD 2.3, *n* = 16), respectively, and did not differ (*P* > 0.57) between the two experimental chambers and air systems.

In the animal experiments, sows were offered half their daily allowance in two modes: a single meal 2 h into the infusion experiment after 12 h of food deprivation (F1), or, 2 to 4 days later, in eight hourly meals starting 2 h before the infusion experiment (F8). One animal refused to eat during the infusions in F1 and F8; these data were discarded, and so all results are for 5 sows. During F1, the fasting ¹⁴CO₂ recovery of 58.1% (SD 4.8) was lower than the mean recovery of 78.8% (SD 5.9) after feeding (*P* = 0.03). In F8, the ¹⁴CO₂ recovery was similar in the CO₂ collection periods when the pigs were offered feed (85.7%, SD 4.4) versus those periods without feeding (78.5%, SD 2.8) (*P* = 0.20); the overall ¹⁴CO₂ recovery during F8 was 82.1% (SD 2.6) which was not different than the recovery after feeding in F1 (*P* = 0.57).

Throughout the entire study period, CO₂ production significantly correlated with ¹⁴CO₂ recovery in F1 (*r* = 0.35, *P* = 0.02) and in F8 (*r* = 0.41, *P* = 0.01). However, this relationship between ¹⁴CO₂ recovery and CO₂ production was not significant in the sub-set of data for F1 immediately after feeding (*P* = 0.30). Figure 1 shows that the CO₂ production in F1 increased sharply immediately after feeding with a subsequent decline to almost fasting values, while the ¹⁴CO₂ recovery remained at a constant high level for 6 h after feeding. In F8, the ¹⁴CO₂ recovery showed little variation throughout the 4 h study (Figure 2).

With respect to minute by minute gas measurements, the CO₂ production of sows fed a single meal (F1, Figure 1) was comparatively low after an overnight fast. Within 10 min after feeding, the CO₂ production almost doubled. This level of CO₂ production was maintained for approximately 27 min, followed by a decrease over a period of about 90 min. Thereafter, the CO₂ production remained fairly constant for 272 min until the end of the experiment. In pigs fed hourly meals (F8, Figure 2), the CO₂ production increased after feeding and remained at a high level for a short period of time (12.5 min, SD 1.7), then declined within approximately 15 min to its initial value and remained there for 29.5 min (SD 3.5) until the next feeding. This pattern was repeated at each feeding. In both F1 and F8, the O₂ consumption and energy expenditure followed the same pattern as the CO₂ production.

In F1, fasting O₂ consumption, CO₂ production and energy expenditure were lower (*P* < 0.01) than immediately after feeding or postprandially (Table 1). The respiratory quotient (RQ) in F1 was greater (*P* = 0.001) postprandially than during fasting, but not different from the peak data. In F8, RQ, O₂ consumption, CO₂ production and energy expenditure were greater (*P* < 0.001) during the peak phase compared to postprandially. CO₂ production and energy expenditure and O₂ consumption during the peak phase were similar (*P* > 0.2) for F1 and F8, whereas for the postprandial phase, these parameters were lower in F1. The RQ was greater (*P* < 0.001) for F1 than for F8 during the peak phase, but similar postprandially. The effect of digestive activity on energy expenditure was 4.54 kJ/min for F1 and 5.66 kJ/min for F8, with a mean of 5.10 kJ/min. The energy expenditure due to physical activity was 11.27 kJ/min for F1 and 9.95 kJ/min for F8, with a mean of 10.61 kJ/min.

Discussion

In this experiment, we studied the effect of feeding regimen on the CO₂ production and ¹⁴CO₂ recovery to support future studies of amino acid kinetics and energy metabolism in pigs and enable comparability with other species. The feeding regimens chosen were: a single meal after an overnight fast to represent the common practice in commercial pig production, and hourly meals as an attempt to approximate a physiological steady state with regard to intended subsequent isotope infusion experiments. Because the absorption of nutrients from casein-based, semi-synthetic diets peaks two hours after a meal [8], the hourly feeding was initiated two hours prior to the start of the CO₂ production and recovery measurements in order to achieve a steady state. Because we did not measure these parameters in the preprandial state of the hourly feeding experiment, the comparison of fasting versus feeding is based on the study involving a single meal, whereas the effect of the feeding regimen is derived from the

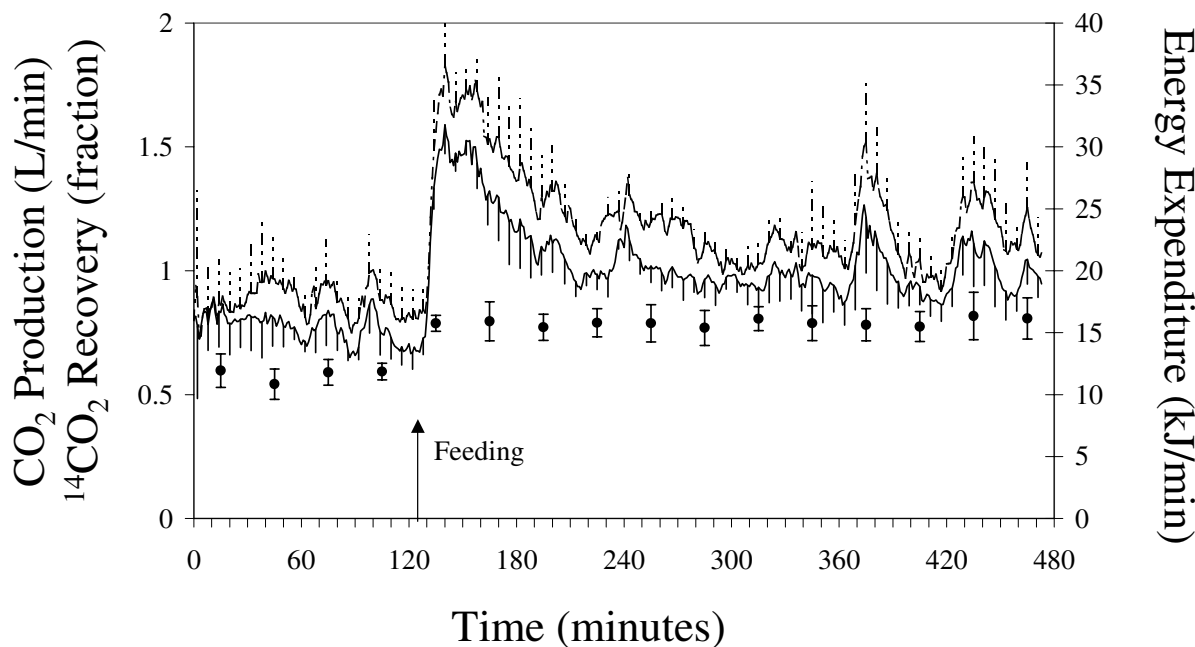


Figure 1
Carbon dioxide production, energy expenditure and $^{14}\text{CO}_2$ recovery in sows given a single meal of half of daily intake. After a 12 h food-deprivation, intravenous $\text{NaH}^{14}\text{CO}_3$ solutions were infused for 2 h prior to feeding and for 6 h after feeding. CO_2 production and energy expenditure were calculated each minute and breath $^{14}\text{CO}_2$ was collected in 30 min periods. Data are means of $n = 5$ sows at each time point. : Energy expenditure. — : CO_2 production. ● : $^{14}\text{CO}_2$ recovery.

comparison of the postprandial state after a single meal versus the entire study period with hourly meals. These comparisons are relevant because the same pigs and the same diets were used for both feeding regimens.

The comparisons of feeding regimens are apparent in the time-course of CO_2 production and energy expenditure (Figs. 1 and 2), which was similar to the time-course of CO_2 production observed by Ball and Bayley [9] in piglets and by el-Khoury et al. [10,11] in humans. In the fasting state, the CO_2 production and energy expenditure was lower than after feeding, consistent with the observations

by el-Khoury et al. [1]. The fasting energy expenditure of 18 kJ/min, or 432 kJ/kg^{0.75}, was close to the maintenance energy expenditure of 458 kJ/kg^{0.75} [5]. Feeding, being a period of intense activity in pigs, caused a sharp increase in the CO_2 production and energy expenditure. The increase in energy expenditure was 10.6 kJ/min or 59% of the fasting energy expenditure and within the range reported by Ramonet et al. [12], who showed that bursts of activity can increase energy expenditure to twice the fasting level. In humans, non-exercise activity thermogenesis has been shown to increase energy expenditure by approximately 40% above the resting metabolic rate [13].

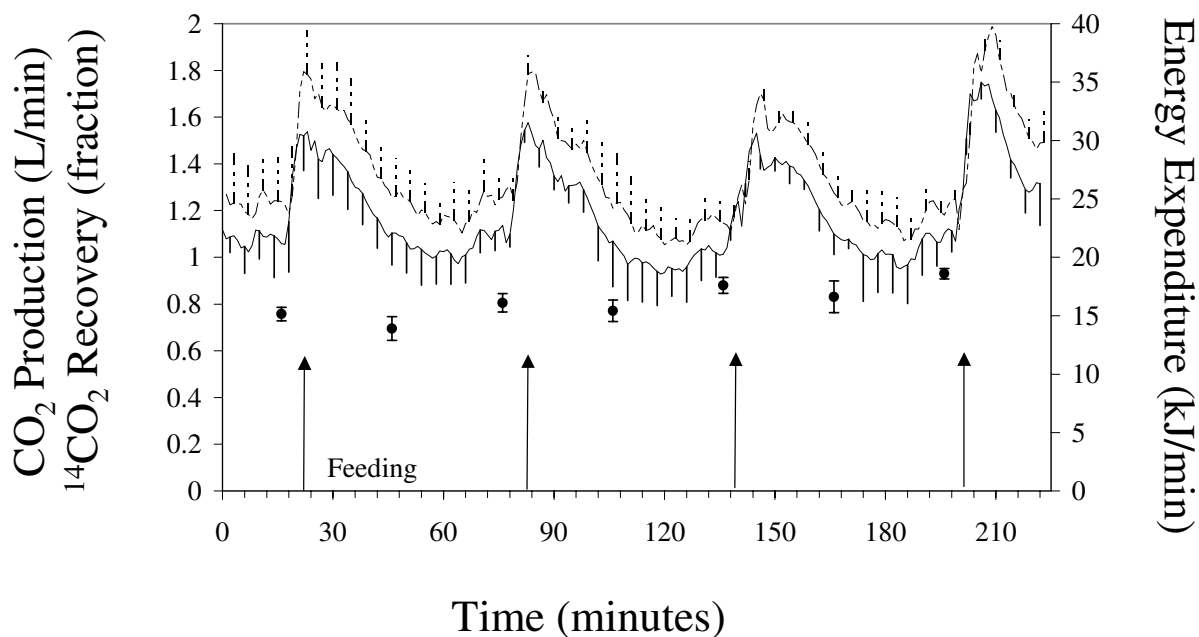


Figure 2
 Carbon dioxide production, energy expenditure and $^{14}\text{CO}_2$ recovery in sows given hourly meals, each of one-eighth of daily intake. After 2 hourly feedings, intravenous $\text{NaH}^{14}\text{CO}_3$ solutions were infused for 4 h with hourly feedings maintained. CO_2 production and energy expenditure were calculated each minute and breath $^{14}\text{CO}_2$ was collected in 30 min periods. Data are means of $n = 5$ sows at each time point. : Energy expenditure. — : CO_2 production. ● : $^{14}\text{CO}_2$ recovery.

Subsequently, the CO_2 production and energy expenditure decreased to a plateau level approximately 29% greater than the fasting CO_2 production and energy expenditure. Because we could not separate the thermal effect of feed from that of activity completely, our estimate was somewhat larger than the estimate of 20% by Ramonet et al. [12]. While the activity-induced energy expenditure was similar for F1 and F8, the postprandial energy expenditure was greater in F8 than F1. This difference may be caused by the presence of the 'short time thermal effect of feeding' [4] which would indicate continuous energy expenditure due to digestion in F8, but not in F1. Despite

this, the average energy expenditure was similar for F1 and F8, probably because the periods of activity were shorter and the decrease in energy expenditure after activity was quicker in F8 than in F1. These data indicate that, similar to humans [14], the comparability of CO_2 production or energy expenditure was maintained between both feeding regimens, although the time-courses were quite different.

However, it appears that the level of CO_2 production and energy expenditure was overestimated in our short time measurement compared to complete 24 h measurements. It can be expected that methane production would

Table 1: Gas exchange parameters of sows receiving either a single (F1) or hourly (F8) meals¹

		Fasting	Peak	Postprandial
CO ₂ (L/min):	F1	0.773 ^a ± 0.052	1.470 ^b ± 0.053	0.990 ^c ± 0.074
	F8	n.d. ²	1.489 ^a ± 0.110	1.039 ^b ± 0.053
	P value, F1 vs F8 ³		0.29	0.001
O ₂ (L/min):	F1	0.871 ^a ± 0.060	1.632 ^b ± 0.056	1.084 ^c ± 0.110
	F8	n.d.	1.614 ^a ± 0.131	1.139 ^b ± 0.062
	P value, F1 vs F8 ³		0.39	0.001
Respiratory quotient (ratio):	F1	0.888 ^a ± 0.029	0.901 ^{ab} ± 0.027	0.916 ^b ± 0.048
	F8	n.d.	0.924 ^a ± 0.039	0.912 ^b ± 0.019
	P value, F1 vs F8 ³		0.003	0.26
Energy expenditure (kJ/min):	F1	17.97 ^a ± 1.21	33.78 ^b ± 1.10	22.51 ^c ± 2.12
	F8	n.d.	33.58 ^a ± 2.60	23.63 ^b ± 1.25
	P value, F1 vs F8 ³		0.64	0.001

¹ Data are means ± SD for *n* = 5 sows. F1 data are from 8 h infusions including a 2 h fasting phase and a single bolus feeding; F8 data are from 4 h infusions with 8 hourly feeds beginning 2 h prior to infusion. For data with letter superscripts within a row, those not sharing a letter are different (*P* < 0.05) across phases and within feeding mode. ² Not determined ³ Differences between feeding modes and within a phase (*P* < 0.05)

amount to about 1% of the digestible energy intake [13]. The energy expenditure was calculated using caloric factors for CO₂ and O₂ (Brouwer formula) [6]. According to this formula, the quantitative excretion via urine of ingested nitrogen would account for a further 0.5% of the digestible energy intake. Therefore, it is unlikely that the omission of methane and urinary energy caused the discrepancy. Rather, the overestimation was probably due to the omission of the respiration measurement during the night when resting animals produce significantly less heat and CO₂. Therefore, the estimates given (kJ/min) should not be extrapolated to 24 h values, unless future experiments show a reliable relationship between short time and 24 h measurements.

The ¹⁴CO₂ recovery correlated well with CO₂ production during both feeding regimens. After an overnight fast, ¹⁴CO₂ recovery (58%) was lower than after feeding either a single meal (79%) or repeated hourly meals (82%). The lower recovery of ¹⁴CO₂ during fasting is consistent with the observations of el-Khoury et al. [1] and Leijssen and Elia [16] in humans. These authors [1] also argued that hourly meals allowed the application of a single recovery factor for isotope infusion studies, based on the lower variability of the ¹⁴CO₂ recovery during frequent feeding. In the present experiment, the variability of the ¹⁴CO₂ recovery data was similar for F1 after feeding and for F8. The mean ¹⁴CO₂ recovery in the postprandial state in our experiment was 81%, which is lower than the 93% reported for parenterally fed piglets [2] and slightly greater than the average recovery of 77% reported for adult humans [1]. In contrast to el-Khoury et al. [1], there appeared to be no strong relationship between ¹⁴CO₂ recovery and CO₂ production immediately after feeding a single meal to sows, probably due to the significantly

greater physical activity in sows versus humans. When fed hourly, however, there was a significant correlation between ¹⁴CO₂ recovery and CO₂ production. In humans, exercise led to an increase in CO₂ recovery [17], although the increase – in accordance with the level of activity – was greater than observed in the sows. The correlation between ¹⁴CO₂ recovery and CO₂ production implies that the CO₂ retention factors adequately reflect the CO₂ production under conditions of hourly feeding.

Conclusions

The CO₂ production or ¹⁴CO₂ recovery was greater after meals than after an overnight fast, but did not differ postprandially between a single meal or hourly meals. Therefore, different retention factors should be applied when studying amino acid oxidation pre- or postprandially. Both feeding regimens led to an overestimation of energy expenditure because the measurements were taken predominantly during the time after feeding and did not incorporate the resting energy expenditure to a sufficient degree. Despite the overestimation, the relative effects of nutritional intervention on energy metabolism can be elicited. To derive valid daily energy expenditure values from a short measurement period, its relationship to a full 24 h measurement period should be established.

The significant correlation of CO₂ production and ¹⁴CO₂ recovery during hourly feeding, but not after a bolus feed, suggests steady state is not achieved in animals after a single feed. This dissociation is probably due to excessive physical activity following the single feeding. Offering hourly meals should therefore be the preferred feeding regimen for future amino acid oxidation studies in animals.

Methods

Equipment

Respiration boxes of about 2 m³ volume were built around commercial farrowing crates and fitted with a feeder and drinker. Air was drawn through these boxes via an inlet at the rear and an outlet above the trough at rates of approximately 240 L/min. After passing through a cold water condenser to remove water from the air, air flow was divided between a series of gas washing bottles for CO₂ collection and a separate air line for gas analysis ('bypass'). The flow rate through each diversion was measured with two commercial air meters (Canadian Meter Corp., Cambridge, Canada). The CO₂ collection stream consisted of five gas washing bottles (four in parallel and a final bottle) followed by a solvent trap. Part of the bypass stream of air was drawn with a small air pump (Gast Model 0531, Gast Mfg. Corp., Benton Harbour, MI) and delivered to oxygen (Taylor-Servomex, Crowborough, UK) and CO₂ analysers (Beckman LB2, Beckman, Irvine, CA). Air flow to the analysers was regulated by ball-type flowmeters (Scienceware Size 2, Fisher Scientific, Mississauga, Canada). The analog output (mV) was converted to digital data by an analog-digital converter (Datagrabber, Data Electronics, Australia) and recorded by a computer. Data acquisition was set for maximum rate (four readings per second) and the average gas concentration for each minute was recorded.

Calibration of respiration systems

The equipment was pressure-tested to ensure that the system contained no air leaks. The recovery of CO₂ and ¹⁴CO₂ by the system was determined by releasing a known quantity of CO₂ and ¹⁴CO₂ in the respiration chambers. Four moles of NaHCO₃, equivalent to the hourly CO₂ production of a 250-kg sow being fed maintenance energy intake, were dissolved in 4 L of water and spiked with 37 kBq of NaH¹⁴CO₃ (American Radiolabeled Chemicals, St. Louis, MO). The solution was placed on a magnetic stirrer inside the respiration chamber and air was collected immediately thereafter. The CO₂ was released by the addition of 4.5 mol of concentrated HCl over 1 h using syringe pumps. After 2 h of air collection in 30 min collection intervals, the quantitative release of CO₂ was confirmed by sampling the bicarbonate solution and measuring the remaining radioactivity. The CO₂ concentration in the bypass flow was recorded at 1 min intervals throughout this 2 h period.

On each study day, the O₂ and CO₂ analysers were calibrated for their zero and gain readings with either pure N₂ (zero) or calibration gas (21% O₂, 1.5% CO₂, 77.5% N₂). The average readings for the zero and calibration gases were each recorded for 5 min immediately before and after each study to correct for any change in analyser readings during the study.

Collection of CO₂

Each gas washing bottle was filled with 100 mL of CO₂ absorber solution (monoethanolamine:2-methoxyethanol, 1:2; Caledon Laboratories, Edmonton, Canada). After each 30 min collection, the pooled volume in the first four bottles and separately, in the fifth bottle, were weighed and sampled. For the first four bottles, 0.5 g of the sample was weighed in a scintillation vial, diluted with 0.5 ml of 2-methoxyethanol, and mixed with 5 mL of a scintillation cocktail (Atomlight, Canberra Packard, Mississauga, Canada); for the fifth bottle, 1 g was weighed and mixed with 5 mL of scintillant. Absorber from the final solvent trap was also weighed, sampled and counted at the end of each study day to verify complete collection. All samples were collected in duplicate and counted in a scintillation counter (Beckman LS 3000, Beckman, Irvine, CA) for 15 min or to a 2 σ error of 2%.

CO₂ recovery in animals: experimental design and diets

Six adult sows (235 kg, SD 5), non-pregnant after their fourth parity, were surgically implanted with catheters in the cephalic vein for isotope infusion. After surgery, they were left to recover for at least 7 d. Between studies, the animals were housed individually in pens 1.5 by 2.1 m fitted with rubber mats. The animal room was temperature controlled at 21 °C. **All procedures were approved by the Faculty Animal Policy and Welfare Committee of the faculty Agriculture, Food and Human Ecology, University of Alberta.**

The semi-synthetic diet contained (g/kg diet) casein (27.5), cornstarch (200.0), corn oil (20.0), sulkaflok (150.0), sugar (527.4), L-cystine (1.67), L-threonine (1.96), L-tryptophan (0.27), L-isoleucine (0.24), L-aspartate (2.02), L-glutamate (2.02), chromic oxide (5.0), limestone (11.0), di-calcium-phosphate (23.0), magnesium sulfate (1.0), potassium bicarbonate (6.2), sodium bicarbonate (3.5), sodium chloride (2.2), thiamin (0.005), vitamin B6 (0.005), vitamin and mineral premix (15.0); the premix contained (per kg) Cu (2000 mg), I (20 mg), Fe (15 g), Mn (1200 mg), Se (30 mg), Zn (12 g), vitamin A (1,000,000 IU), vitamin D (100,000 IU), vitamin E (8000 IU), vitamin K (200 mg), biotin (20 mg), choline (100,000 mg), folacin (160 mg), niacin (4000 mg), pantothenic acid (2500 mg), riboflavin (1200 mg), and vitamin B₁₂ (3000 μ g). This diet provided twice the maintenance requirement for amino acids and at least 120% of the mineral and vitamin recommendations of NRC [5] for gestating sows. The diets were offered to exceed the maintenance energy requirement [5] by 10%. Between study days, the animals were fed twice daily. On study days, sows were offered half their daily allowance in two modes: a single meal 2 h into the infusion experiment after 12 h of food deprivation (F1), or, 2 to 4 days later, in eight hourly meals starting 2 h before the infusion exper-

iment (F8). The hourly feeding regimen was imposed on the day of study only. Each animal was subjected to both treatments in a randomized crossover manner.

Isotope infusion

The recovery of $^{14}\text{CO}_2$ was determined during primed, constant infusions of a saline solution containing $\text{NaH}^{14}\text{CO}_3$. The priming dose of 20% of the hourly infusion dose of $\text{NaH}^{14}\text{CO}_3$ was administered within 1 min. The constant dose was infused by syringe pump (Fisher Scientific, Mississauga, Canada) at an approximate rate of 95 kBq/h for either 8 h (F1) or 4 h (F8); the infusion solution was weighed and sampled immediately after each infusion to measure actual radioactivity delivered. Throughout the study, $^{14}\text{CO}_2$ was collected in 30-min intervals. Because repeated measurements were conducted in each animal, the background radioactivity in breath was determined during a 30 min CO_2 collection before each isotope infusion. This background collection was started as soon as the CO_2 content in the air drawn from the box had reached an equilibrium as measured by the CO_2 analysers. Oxygen consumption and CO_2 production were monitored in 1 min average measurements throughout the infusion studies.

Calculations

The air flow in the respiration system was corrected to 0°C and an atmospheric pressure of 760 mm Hg. Oxygen consumption and CO_2 production were calculated by multiplying the corrected air flow by the 1 min average gas concentration after correcting for room air. The respiratory quotient (RQ) was calculated by dividing the CO_2 production by the O_2 consumption. The energy expenditure was calculated using caloric factors for CO_2 and O_2 (Brouwer formula) [6]. Methane production and urinary N excretion were omitted from the formula because they were not measured. The CO_2 production was divided into three phases: 1) the fasting phase in F1 prior to feeding, 2) the period of high CO_2 production immediately after each feeding in both F1 and F8, and 3) the postprandial phase following this period of high CO_2 production in both F1 and F8. A non-significant regression of CO_2 production on time was used to define plateaus during these phases. The resulting time periods were used to estimate the O_2 consumption, RQ and heat production during the three phases. The effect of digestion was calculated by subtracting the fasting data from the postprandial data. The effect of physical activity was estimated by subtracting the data from the postprandial phase from the data directly after feeding. To compare the CO_2 production to the $^{14}\text{CO}_2$ recovery, 30-min values were calculated for the CO_2 production to coincide with the collection of $^{14}\text{CO}_2$.

Statistics

Data are presented as means and SD. One-minute means were calculated within feeding frequencies and phases assuming 'sow' to be a random effect. Data across three phases within F1 feeding were analyzed by ANOVA using Fisher's (protected LSD) multiple comparisons between groups and were considered significant when $P < 0.05$ [7]. Differences between the two phases of F8 feeding as well as the effects of the feeding regimen on CO_2 excretion, O_2 consumption, $^{14}\text{CO}_2$ recovery, and RQ were evaluated using Student's t-test. The relationship between CO_2 production and $^{14}\text{CO}_2$ recovery was assessed using Pearson's correlation coefficients.

Authors' contributions

SM and RFPB designed the experiment, carried out the animal studies, analyzed the data and drafted the manuscript. PBP and ROB supervised the overall project and edited the manuscript. All authors read and approved the final manuscript.

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References

1. el-Khoury AE, Sánchez M, Fukagawa NK, Gleason RE, Young VR: **Similar 24-h pattern and rate of carbon dioxide production, by indirect calorimetry vs. stable isotope dilution, in healthy adults under standardized metabolic conditions.** *J Nutr* 1994, **124**:1615-1627.
2. Wykes LJ, House JD, Ball RO, Pencharz PB: **Amino acid profile and aromatic amino acid concentration in total parenteral nutrition: effect on growth, protein metabolism and aromatic amino acid metabolism in the neonatal piglet.** *Clin Sci (Lond)* 1994, **87**:75-84.
3. Tabiri HY, Bertolo RFP, Ball RO, Korver DR: **Development of the indicator amino acid oxidation technique in chickens: Calibration of oxidation system and determination of bicarbonate retention factor.** *Poult Sci* 2002, **81**:1020-1025.
4. Van Milgen J, Noblet J: **Modelling energy expenditure in pigs.** In: *Modelling Nutrient Utilization in Farm Animals* Edited by: McNamara JP, France J, Beever DE. New York, NY, CAB International; 2000:103-114.
5. National Research Council (NRC): *Nutrient Requirements of Swine* 10th edition. Washington, DC, National Academy Press; 1998.
6. McLean JA, Tobin G: *Animal and human calorimetry* Cambridge, UK, Cambridge University Press; 1987.
7. Statistical Analysis System (SAS): *SAS/Stat User Guide, Release 6.03 Edition* Cary, NC, SAS Institute; 1988.
8. Darcy-Vrillon B, Vaugalade P, Bernard F, Duée PH: **Effect of food intake level on arterial glycaemia, portal glucose appearance and intestinal oxygen consumption in the pig.** *Sciences des Aliments* 1999, **19**:367-375.
9. Ball RO, Bayley HS: **Time course of the total and radioactive carbon dioxide production by piglets receiving dietary [^{14}C]phenylalanine.** *Can J Physiol Pharmacol* 1985, **63**:1170-1174.
10. el-Khoury AE, Fukagawa NK, Sánchez M, Tsay RH, Gleason RE, Chapman TE, Young VR: **Validation of the tracer-balance concept with reference to leucine: 24-h intravenous tracer studies with L-[^{1-13}C]leucine and [^{15}N - ^{15}N]urea.** *Am J Clin Nutr* 1994, **59**:1000-1011.
11. el-Khoury AE, Sánchez M, Fukagawa NK, Gleason RE, Tsay RH, Young VR: **The 24-h pattern kinetics of leucine oxidation in healthy adults receiving a generous leucine intake via three discrete meals.** *Am J Clin Nutr* 1995, **62**:579-590.

12. Ramonet Y, van Milgen J, Dourmad JY, Dubois S, Meunier-Salaun MC, Noblet J: **The effect of dietary fibre on energy utilisation and partitioning of heat production over pregnancy in sows.** *Br J Nutr* 2000, **84**:85-94.
13. Levine J, Melanson EL, Westterterp KR, Hill JO: **Measurement of the components of nonexercise thermogenesis.** *Am J Physiol Endocrinol Metab* 2001, **281**:E670-E675.
14. Verboeket van der Venne WPHG, Westterterp KR: **Influence of the feeding frequency on nutrient utilization in man: Consequences for energy metabolism.** *Europ J Clin Nutr* 1991, **45**:161-169.
15. Noblet J, Shi XS: **Comparative digestibility of energy and nutrients in growing pigs fed ad libitum and adult sows fed at maintenance.** *Livest Prod Sci* 1993, **34**:137-152.
16. Leijssen DP, Elia M: **Recovery of $^{13}\text{CO}_2$ and $^{14}\text{CO}_2$ in human bicarbonate studies: a critical review with original data.** *Clin Sci (Lond)* 1996, **91**:665-677.
17. Trimmer JK, Casazza GA, Horning MA, Brooks GA: **Recovery of $^{13}\text{CO}_2$ during rest and exercise after $[1-(^{13}\text{C})\text{acetate}$, $[2-(^{13}\text{C})\text{acetate}$ and $\text{NaH}^{(13)}\text{CO}_3$ infusions.** *Am J Physiol Endocrinol Metab* 2001, **281**:E683-E692.

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