

Diet and trophic position of Leach's storm-petrel *Oceanodroma leucorhoa* during breeding and moult, inferred from stable isotope analysis of feathers

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ABSTRACT: We combined conventional dietary sampling (2002) with stable isotope analysis of nitrogen ($\delta^{15}\text{N}$) and carbon ($\delta^{13}\text{C}$) in feathers of Leach's storm-petrels *Oceanodroma leucorhoa* (2001 and 2002) to investigate temporal, sex- and age-related variation in diet and trophic position, at Baccalieu Island, Newfoundland, Canada. Nestlings are fed mainly fish and crustaceans (ca. 90 and 9% by mass, respectively) toward the end of chick-rearing. The bulk of the fish were vertically migrating myctophids, associated with the offshore environment. Crustaceans were mainly parasitic hyperiid amphipods *Hyperia galba*, commonly found nearshore in association with scyphozoan jellyfish hosts. $\delta^{15}\text{N}$ values in summer-grown feathers were similar between years, but differed between age-classes in 2001 when chicks occupied slightly higher trophic level (3.2 vs. 3.1). $\delta^{13}\text{C}$ values in summer-grown feathers were lower for chicks than adults. Predictions from a 3-source dual-isotope linear mixing model indicated that adults consumed 47% glacier lanternfish *Benthoosema glaciale*, 5% capelin *Mallolotus villosus* and 49% *H. galba* in 2001, while in 2002 respective values were 87, 6 and 7%. Model output for chicks was not biologically meaningful, possibly due to the lipid-rich (and $\delta^{13}\text{C}$ depleted) stomach oil fed to chicks by adults. For adults, no sex differences were found in values of either isotope during summer. $\delta^{15}\text{N}$ values were significantly higher in feathers grown during summer than during mid-moult, suggesting increased consumption of crustaceans in winter. $\delta^{13}\text{C}$ values in adult primaries were relatively enriched, consistent with movement of post-breeding birds toward tropical waters. Combining conventional and chemical analyses has provided some of the first insights into the year-round ecology of the smallest and most abundant seabird breeding in eastern North America.

KEY WORDS: Stable-isotopic analysis · Trophic level · Diet · Moult · *Oceanodroma leucorhoa* · Newfoundland · Storm-petrel

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INTRODUCTION

The food and feeding ecology of seabirds in general, and Procellariiformes in particular, has been relatively well studied during summer when birds return to land to breed. In contrast, the non-breeding period that often extends well over half of the year is poorly studied as most species spend their time exclusively at sea. While archival and telemetric devices are revolutioniz-

ing information on feeding ecologies—foraging and migratory strategies of larger species both during and outside breeding (e.g. Weimerskirch et al. 1993, 2002, Croxall et al. 2005, Garthe et al. 2006)—the lives of species too small to carry such devices remain relatively elusive.

Quantification of diets during breeding has traditionally been based on analysis of stomach content, but stable isotope analysis (SIA) of various tissues has also

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been used to estimate diet and trophic relationships for a variety of seabird communities (e.g. Hobson 1993, Hobson et al. 1994, Hodum & Hobson 2000). SIA has also recently been used to identify foraging locations and therefore movements of birds between breeding and non-breeding seasons (e.g. Hobson 1999, Cherel et al. 2000, Quillfeldt et al. 2005). Isotope ratio variation in bird tissue reflects that in the local environment and is determined by the relative proportions of isotopically distinct dietary components (Hobson 1993). Feathers are an attractive tissue to sample because they can be collected non-destructively and feather keratin is metabolically inert after synthesis, so isotopic composition reflects diet at the time feathers were grown (e.g. Mizutani et al. 1992). For many procellariiform seabirds, breeding and moult are essentially exclusive in time, so determining the isotopic composition of feathers grown at different times of year represents a method of gathering information on temporal variation in feeding ecology (Cherel et al. 2000, Quillfeldt et al. 2005).

We investigated temporal, sex- and age- (adult vs. chick) related variation in the diet and trophic position of Leach's storm-petrel *Oceanodroma leucorhoa* at the center of its breeding range in the northwest Atlantic by combining conventional diet sampling with analyses of stable nitrogen and carbon isotope ratios in feathers of adults and chicks. Leach's storm-petrel is the smallest (ca. 50 g) and most abundant seabird breeding in eastern North America. It is highly pelagic, feeding up to many hundreds of km from the colony, beyond the edge of the continental shelf (Brown 1986, G. K. Davoren et al. unpubl. data), and it feeds at the ocean surface. Adult-chick comparisons were made during the 2001 and 2002 breeding seasons using stable isotope values in feathers grown during the previous weeks at the breeding colony. Adult Leach's storm-petrels begin moulting tail feathers on the breeding grounds in August, some 4 to 6 wk before chicks begin fledging (A. Hedd pers. obs.). During 2002, we also investigated chick diet using conventional sampling techniques. SIA was used to investigate relative changes in the trophic position of adults between breeding (summer) and moult (winter). Adults depart the colony after chicks fledge and are reported to winter in the tropics (Cramp & Simmons 1977, Huntington et al. 1996). We contrasted adult tail feathers, grown on the breeding grounds, with a sequence of primary feathers (P1, P4, P7, P10) grown during the previous winter moult. Adult primary moult commences after birds depart the colony in fall and is complete before they return to breed the next season (Huntington et al. 1996). Combining conventional diet sampling with SIA allowed us to investigate temporal and age-related variation in diet and feeding ecology of Leach's storm-petrel through its annual cycle.

MATERIALS AND METHODS

Species, study site and field collections. This study was conducted in the Baccalieu Island Ecological Reserve (48.12° N, 52.80° W), off the east coast of Newfoundland, Canada, where it is estimated that 3.5 million pairs of Leach's storm-petrels breed annually (Sklepkovych & Montevecchi 1989). Within our study area, egg-laying begins late in May, the mean chick hatch date approximates late July and fledging begins mid-September (Huntington et al. 1996, A. Hedd unpubl. data). As part of another study (M. Logan unpubl. data), 10 adult-chick pairs (11 adults and 10 chicks) of Leach's storm-petrels were collected during late chick-rearing (early September) 2001, and a further 15 pairs (16 adults and 15 chicks) were collected in early September 2002 (Canadian Wildlife Service Scientific Take Permits SC2207 and SC2249). Birds were frozen in the field and maintained at -20°C with the exception of brief thaws for tissue extraction, feather sampling and gender determination via dissection (adults only).

We obtained feather samples from these carcasses. Adults moult their tail and primary feathers annually (Cramp & Simmons 1977). Tail moult begins late in the breeding season, while primary moult occurs solely outside the breeding period (Ainley et al. 1976, Cramp & Simmons 1977). Primary moult is descendant, beginning October-November when birds may still be near the breeding grounds, and continuing during migration and on the tropical wintering grounds (Ainley et al. 1976, Cramp & Simmons 1977, Hémery & Jouanin 1988). Precise timing of primary moult is unknown but it is relatively protracted. Hémery & Jouanin (1988) reported that all Age 1+ birds recovered during a mid-December wreck in the Bay of Biscay were moulting primary feathers. The outer 1 to 3 feathers were well worn (old), the next 1 to 2 were falling out or growing, while the remaining inner feathers had already been replaced. Primary moult is completed before the start of the next breeding season. In each year, we consequently sampled adult tail feathers grown at the breeding colony (2001 and 2002) and a sequence of primary feathers (from the right wing) grown at the beginning (P1), middle (P4, P7) or end (P10) of the 2000 or 2001 winter moult. For 6 adults (1 in 2001 and 5 in 2002) we used growing body feathers as opposed to tail feathers for the breeding season sample as these birds had just commenced tail moult and the tail feathers did not yield adequate material. For each chick, we sampled the tip of the most recently grown primary on the right wing. For the majority of chicks this was P10, but lower primaries were used for younger chicks. We also obtained specimens of 2 of the main prey types (myctophid or lanternfishes and Hyperiid amphipods)

delivered to Leach's storm-petrel chicks in Newfoundland colonies (authors' unpubl. data). The glacier lanternfish *Benthoosema glaciale* were collected on board CCGS 'Teleost' during standard capelin *Malloctus villosus* tows (using 12 mm mesh nets) in May 2002, while jellyfish *Aurelia aurita* and *Cyanea capillata* were collected with pole nets from a small vessel outside St. John's harbour, Newfoundland, in September 2002. Hyperiid amphipods *Hyperia galba* were removed from the jellyfish gonads. Prey samples fell within the size range consumed by Leach's storm-petrels, and they were stored frozen until analysis.

Between 31 August and 1 September 2002, 49 diet samples were collected from adults caught in mist-nets at night. Plastic tarps laid under nets facilitated sample recovery as many birds spontaneously regurgitated stomach contents either upon impact with or when being removed from the net. Birds were subsequently inverted and their stomachs gently massaged to obtain any food that remained in the gut. Samples were placed in individual bags and frozen. In the laboratory, samples were thawed, and weighed before and after draining to estimate the mass contribution of liquid and solid material. Solid material was subsequently washed through a series of stacked sieves, placed in a sorting tray and separated into broad prey classes (fish, cephalopods, crustaceans, others). Each prey class was weighed to estimate the fresh mass contribution. Within each class, individual prey were identified to the lowest possible taxon and counted. Samples were generally well digested, and all fish were identified from otoliths using Härkönen (1986), Campana (2004) and material held at the Department of Fisheries and Oceans in St. John's. Crustaceans were identified following Dunbar (1963). Otoliths were paired, counted separately left and right and the number of fish was estimated from the larger count. When fish was noted from flesh or bones alone, we assumed it represented 1 individual. Total length of intact amphipods and euphausiids was measured, under a dissecting microscope fitted with a graticule, from the front of the eye to the tip of the uropods and from the tip of the rostrum to the tip of the uropods, respectively. Body length of fish was estimated from uneroded otolith dimensions using either published relationships (Härkönen 1986, Winters 1989, Olsson & North 1997) or our own equations. To reconstruct the mass composition of the diet, the body masses of crustaceans and fish were estimated from lengths using either published relationships (Härkönen 1986), or our own equations. Species-specific equations were used where possible, but when not available, estimates were made from closely related species or from species with similar morphology. The reconstructed mass of each species/taxon was calculated by multiplying average body mass by the

number of individuals recovered. The calculated mass for all taxa was pooled and expressed as a percentage of the total. In total, 41% of fish (52 of 128) were unidentified and to conduct the mass reconstruction, we assumed that these fish were distributed (by number) in the same relative proportion as those we identified. Data are presented as percent frequency of occurrence, number and reconstructed mass.

Tissue preparation, stable isotope measurements and models. Feathers were placed in glass scintillation vials and cleaned of surface contaminants using a 2:1 ratio solution of chloroform:methanol. Feathers were air dried for 24 h and cut into small fragments using stainless steel scissors. We avoided the quill and shaft. For fish, a piece of muscle was removed from each thawed specimen, while a number of whole amphipods (≥ 10) were combined to create a single sample. Prey samples were oven dried to constant mass at $+60^{\circ}\text{C}$, coarsely ground and lipids were removed using a 2:1 ratio solution of chloroform:methanol. Amphipods were additionally placed in 1 N HCl to remove carbonates (for ca. 1 h), and then rinsed with distilled water (for ca. 2 h). Prey samples were then ground to a fine powder using a mortar and pestle. From each feather and prey sample, a 1 mg subsample was weighed into a tin cup that was crushed into a pellet and saved for isotopic analysis. All instruments were cleaned with acetone between samples to prevent cross-contamination. Samples were sent to David Harris at the Stable Isotope Facility, University of California, Davis, USA, for determination of isotope values.

Stable isotope values are expressed in delta notation (δ) as parts per thousand (‰) according to the following:

$$\delta X = [(R_{\text{sample}}/R_{\text{standard}}) - 1] \times 1000 \quad (1)$$

where X is ^{15}N or ^{13}C and R is the corresponding ratio $^{15}\text{N}/^{14}\text{N}$ or $^{13}\text{C}/^{12}\text{C}$. R_{standard} for ^{15}N and ^{13}C are atmospheric N_2 (AIR) and the PeeDee Belemnite (PDB) standard, respectively. Replicate measurement of laboratory standards (2 standards for every 12 unknowns) indicated measurement errors of approximately 0.2 and 0.1‰ for nitrogen and carbon, respectively.

While carbon is not generally used to indicate trophic position (Hobson & Welch 1992), we simultaneously determined carbon and nitrogen in our samples. Two isotopes are superior at segregating species and presumably distinguishing temporal differences within a species, and stable carbon isotope ratios have previously been used to infer inshore or benthic vs. offshore or pelagic feeding in seabirds (e.g. Hobson et al. 1994). Isotope values of food fractionate or change when incorporated into consumer tissues according to the following:

$$D_t = D_d + \Delta_{dt} \quad (2)$$

where D_t is isotopic value in the tissue, D_d is isotopic value in the diet, and Δ_{dt} is isotopic fractionation factor between tissue and diet. Cherel et al. (2005) reviewed all control studies and reported average discrimination factors between lipid-extracted whole prey and feathers for fish-eating birds at $+4.2 \pm 0.7\text{‰}$ ($n = 12$) for $\delta^{15}\text{N}$ and $+1.2 \pm 1.2\text{‰}$ ($n = 6$) for $\delta^{13}\text{C}$, and we adopted their values. Stable nitrogen and stable carbon values of fish (herring *Clupea harengus* and capelin) were also found to differ depending on whether they derived from muscle tissue or whole specimens (Cherel et al. 2005), with whole fish being depleted in $\delta^{15}\text{N}$ by $-0.9 \pm 0.1\text{‰}$ and enriched in $\delta^{13}\text{C}$ by $+0.3 \pm 0.1\text{‰}$. As both our values for fish and those from the literature (Sherwood & Rose 2005) were derived from muscle, they were appropriately adjusted prior to applying models to estimate dietary contributions. We used the 3-source dual-isotope mixing model (Phillips & Gregg 2001) which provides standard errors and confidence intervals for source proportion estimates, accounting for the variability observed in sources (prey species) as well as the mixture (feathers). The main prey of chick-rearing Leach's storm-petrel in Newfoundland are myctophid fishes (represented by the glacier lanternfish) and hyperiid amphipods *Hyperia galba* (authors' unpubl. data), with capelin making a contribution at some sites in some years. We estimate that lanternfishes, hyperiid amphipods and capelin together constituted 87% of chick diet at Baccalieu in 2002.

To calculate the trophic level (TL) of a consumer (TL_{consumer}), we used the mean TL for pelagic zooplankton on the Newfoundland and Labrador continental shelf (2.1; Sherwood & Rose 2005) and an isotopic enrichment factor between TLs of 3.6‰, calculated by Fry (1988) for the George's Bank region. The TL in the food web may be calculated as:

$$TL_{\text{consumer}} = 2.1 + (D_{\text{consumer}} - D_{\text{pelagic zooplankton}})/3.6 \quad (3)$$

where D_{consumer} is the isotopic signature of the consumer and $D_{\text{pelagic zooplankton}}$ is that for pelagic zooplankton (9.9‰, Sherwood & Rose 2005).

Effects of age (adult vs. chick) and sex on isotope values in feathers grown during the breeding period, and interactions between factors, were tested with 1- or 2-

Table 1. *Oceanodroma leucorhoa*. Stable nitrogen ($\delta^{15}\text{N}$) and carbon isotope ($\delta^{13}\text{C}$) values (‰, mean \pm SD) in feathers of Leach's storm-petrel from Baccalieu Island, Newfoundland, Canada, as a function of age, year and stage of the annual cycle. Winter samples consist of primary feathers grown at the beginning (P1), middle (P4, P7) or end (P10) of the moult. Trophic levels (TL) were estimated using Eq. (3)

Age (feather type)	n	Year	$\delta^{15}\text{N}$	$\delta^{13}\text{C}$	TL
Breeding (chick rearing)					
Chicks (primaries)	10	2001	14.0 \pm 0.5	-19.3 \pm 0.3	3.2 \pm 0.1
	15	2002	13.6 \pm 0.7	-19.5 \pm 0.4	3.1 \pm 0.2
Adults (Tail)	11	2001	13.4 \pm 0.4	-18.8 \pm 0.6	3.1 \pm 0.1
	16	2002	13.6 \pm 0.7	-18.2 \pm 0.6	3.1 \pm 0.2
Winter/moulting					
Adults (P 1)	11	2000/01	13.4 \pm 0.7	-18.2 \pm 0.4	3.1 \pm 0.2
	16	2001/02	13.2 \pm 0.9	-17.7 \pm 0.7	3.0 \pm 0.3
Adults (P 4)	11	2000/01	12.7 \pm 1.2	-18.1 \pm 0.8	2.9 \pm 0.3
	16	2001/02	12.7 \pm 1.0	-17.6 \pm 0.6	2.9 \pm 0.3
Adults (P 7)	11	2000/01	12.4 \pm 1.0	-17.9 \pm 0.7	2.8 \pm 0.3
	16	2001/02	13.1 \pm 1.2	-17.4 \pm 0.8	3.0 \pm 0.3
Adults (P 10)	11	2000/01	12.7 \pm 1.3	-17.4 \pm 0.4	2.9 \pm 0.3
	16	2001/02	13.1 \pm 1.2	-17.4 \pm 0.7	3.0 \pm 0.3

Table 2. Stable nitrogen ($\delta^{15}\text{N}$) and carbon isotope ($\delta^{13}\text{C}$) values (‰, mean \pm SD) in lipid-free fish muscle and whole amphipods collected off eastern Newfoundland, Canada, in 2002. Trophic levels (TL) were estimated using Eq. (3). SL: standard length

Prey species	n	SL (mm)	$\delta^{15}\text{N}$	$\delta^{13}\text{C}$	TL
Glacier lanternfish	10	57 \pm 5	10.2 \pm 0.5	-19.5 \pm 0.6	2.2 \pm 0.1
Hyperiid amphipod	8	-	8.9 \pm 0.5	-20.7 \pm 0.6	1.8 \pm 0.1

way ANOVA, as appropriate. Effects of year, sex, and time of collection on isotope values in adult feathers, and interactions between factors, were tested using repeated measures ANOVA with the Wilk's Lambda statistic. When significant differences ($p < 0.05$) were found, pairwise comparisons were conducted without correction for multiple comparisons. All tests except Levene's test for equal variance (Minitab release 13.30) were conducted using Systat for Windows (v10.2). Values are presented as means \pm 1 SD.

RESULTS

Stable isotope analysis

$\delta^{15}\text{N}$ values ranged from 8.9 \pm 0.5‰ in *Hyperia galba* to 14.0 \pm 0.4‰ in feathers of Leach's storm-petrel chicks in 2001 (Tables 1 & 2, Fig. 1). $\delta^{13}\text{C}$ values ranged from -20.7 \pm 0.6‰ in *H. galba* to -17.4 \pm 0.4‰ in adult primary feathers (P10) grown in the 2000/01 inter-

breeding (moulting) period. During the breeding season, feather $\delta^{15}\text{N}$ values were similar between years for adults (2-way ANOVA, $F_{1,23} = 0.68$, $p = 0.42$) and for chicks ($F_{1,23} = 1.78$, $p = 0.20$), but differed between age classes in 2001 when values for chicks were higher ($F_{1,19} = 10.22$, $p = 0.005$). In 2002, $\delta^{15}\text{N}$ values were similar for adults and chicks ($F_{1,29} = 0.01$, $p = 0.92$). $\delta^{13}\text{C}$ values were similar between years for chicks ($F_{1,23} = 3.09$, $p = 0.092$), whereas for adults, values were significantly higher in 2002 (2-way ANOVA $F_{1,23} = 8.27$, $p = 0.009$). $\delta^{13}\text{C}$ values differed between these age-classes, being significantly higher in adult than in chick feathers in 2002 ($F_{1,29} = 53.37$, $p < 0.001$); a non-significant trend that was also observed in 2001 ($F_{1,19} = 3.94$, $p = 0.062$). There were no significant sex effects or sex by year interactions for values of either isotope in adult feathers grown during the breeding season.

$\delta^{15}\text{N}$ values varied significantly among adult primary feathers (2-way repeated measures ANOVA, Wilk's Lambda $F_{3,21} = 5.12$, $p = 0.008$), with those for P4 and P7 (grown during mid-moult), being significantly lower than for P1, which is the first flight feather to be renewed ($p < 0.05$). There were no effects of sex or year on $\delta^{15}\text{N}$ values in these feathers. For $\delta^{13}\text{C}$, there was significant between-year variation (2-way repeated measures ANOVA, Wilk's Lambda $F_{1,23} = 6.79$, $p = 0.016$), so the years were examined separately. In 2001, $\delta^{13}\text{C}$ values varied significantly among adult primary feathers (repeated measures ANOVA, Wilk's Lambda $F_{3,8} = 6.73$, $p = 0.014$), being significantly greater in P10 than in P1, P4 or P7 ($p < 0.05$). $\delta^{13}\text{C}$ values were similar in all primary feathers grown during the 2001/02 moult (repeated measures ANOVA, Wilk's Lambda $F_{3,13} = 0.55$, $p = 0.660$).

For adults, we also contrasted feathers grown in summer and winter. $\delta^{15}\text{N}$ values varied significantly (2-way repeated measures ANOVA, Wilk's Lambda $F_{4,20} = 9.98$, $p < 0.001$), and were higher in feathers grown at the colony in summer than in those grown during mid-moult in winter (P4 and P7; $p < 0.01$). This tendency was also observed for the final primary (P10), but the result was marginally non-significant ($p = 0.051$). In both years, $\delta^{13}\text{C}$ values were significantly different between breeding and moult (repeated measures ANOVA, Wilk's Lambda, for 2001 $F_{4,7} = 7.81$, $p = 0.10$, and for 2002

$F_{4,12} = 4.98$, $p = 0.013$), and in both years, values in feathers grown on the breeding grounds were significantly lower than in all feathers grown during winter ($p < 0.05$ for all).

We used variance in $\delta^{15}\text{N}$ data as a proxy for dietary breadth (Hodum & Hobson 2000), and examined the relative degree of individual variability during breeding and moult. Mean $\delta^{15}\text{N}$ variance across years for adults during breeding ($0.36 \pm 0.29\%$) was significantly lower than observed during moult (Levene's test = 10.31, $p < 0.01$; P1 = $0.66 \pm 0.24\%$, P4 = $1.21 \pm 0.16\%$, P7 = $1.14 \pm 0.31\%$, P10 = $1.55 \pm 0.05\%$). Chick dietary breadth was also relatively low ($0.37 \pm 0.22\%$) and showed similar variability to that of adults (Levene's test = 0.66, $p = 0.42$).

Food samples

Food samples ($n = 49$) collected from adult storm-petrels returning to Baccalieu Island, Newfoundland, on 31 August to 1 September 2002 averaged 5.2 ± 2.5 g

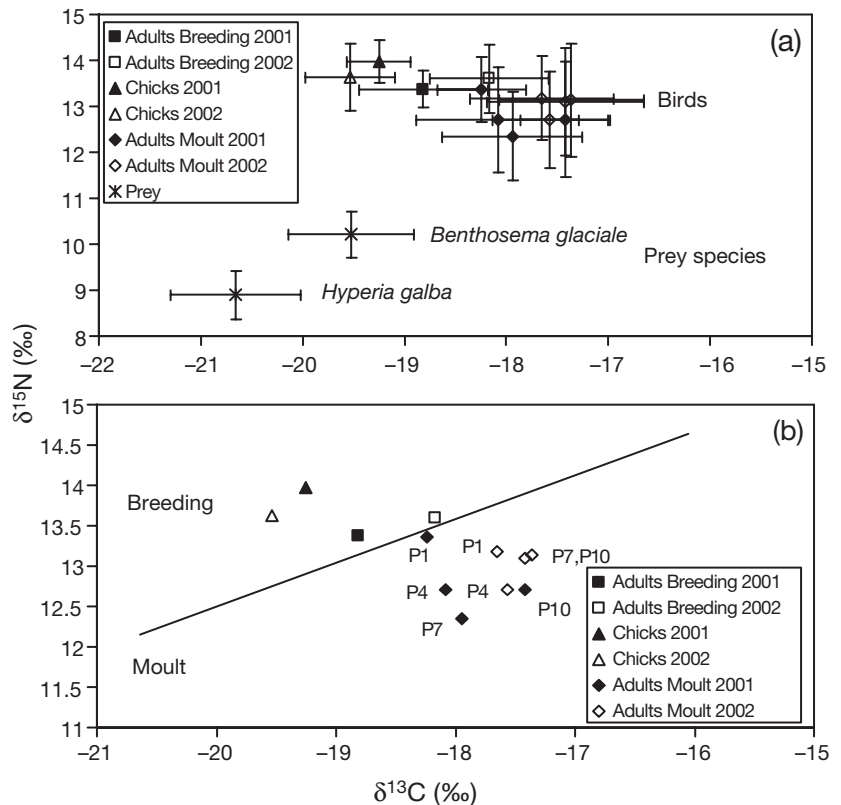


Fig. 1. *Oceanodroma leucorhoa*. Mean (\pm SD) $\delta^{15}\text{N}$ and carbon $\delta^{13}\text{C}$ values of (a) Leach's storm-petrels and their prey in 2001 and 2002. (b) Values for bird only measurements (error bars omitted for clarity). Diagonal line separates the breeding and moulting periods. For adults, moulting samples consist of primary feathers grown at the beginning (P1), middle (P4, P7) or end (P10) of winter

(range 0.3 to 10.9 g). By mass, these samples were 57 % solid and 43 % liquid. The solid portion (n = 48) was composed of 80.3 % fish, 19.7 % crustacean and <0.1 % cephalopod. Fish dominated (i.e. >50 % by fresh mass) in 43 of the samples, while crustaceans dominated in 5 samples.

Chick food was dominated by fish, which accounted for 90.0% by reconstructed mass (Table 3). Crustaceans were the only other important prey class, accounting for 9.2% of diet mass, while remains of a single squid contributed the rest (0.7%). A total of 2264 prey were recovered, including 2136 (94.3%) crustaceans, 128 (5.7%) fishes and 1 (<0.1%) cephalopod. The fish component of the diet was generally well digested, being dominated by offshore species from

the family Myctophidae (66 of 76 identified fish). As only 59% (76 of 128) of fish could be identified, 37% of the overall diet mass was placed in an unidentified/digested fish category (Table 3). Myctophids accounted for 77% of the mass of identified fish (41.2 of 53.4%; Table 3) and sexually mature individuals of 2 species were important: the glacier lanternfish and Arctic telescope *Protomyctophum arcticum*. Three spotted lanternfish *Myctophum punctatum* were also recovered. Capelin and sand lance *Ammodytes* spp. were also important to storm-petrel chicks in 2002, contributing 6.6 and 3.7% by mass, respectively. Capelin and sandlance each occurred in just 6% of samples, but the large size of capelin in particular (7.5 g) resulted in its significant mass contribution. The

Table 3. *Oceanodroma leucorhoa*. Chick diet of Leach's storm-petrel (n = 49) at Baccalieu Island, Newfoundland, 31 August to 1 September 2002. % RM: reconstructed mass

	Occurrence		%RM	Body length (mm)		n	
	%	n		% n	Mean (SD)		Range
Fish	91.8	128	5.7	90.0			
Myctophidae							
<i>Benthoosema glaciale</i>	38.8	23	1.0	14.5	57.1 (6.2)	46.9–67.9	19
<i>Ceratoscopelus maderensis</i>	2.0	1	<0.1	0.6			
<i>Protomyctophum arcticum</i>	26.5	20	0.9	12.3	67.2 (3.1)	63.0–70.9	8
<i>Myctophum punctatum</i>	6.1	3	0.1	1.9			
<i>Lampanyctus</i> sp. ?	2.0	1	<0.1	0.6			
Digested myctophids	28.6	18	0.8	11.3			
Gadidae							
Hake or <i>Micromesistius</i>	2.0	1	<0.1	1.1	3,59		1
Osmeridae							
<i>Mallotus villosus</i>	6.1	3	0.1	6.6	104.5 (9.4)	93.7–110.5	3
Ammodytidae							
<i>Ammodytes</i> sp.	6.1	5	0.2	3.7	90.0 (38.1)	53.5–129.5	3
Scorpaeniformes	2.0	1	<0.1	0.8			
Unidentified/digested fish	59.2	52	2.3	36.6			
Crustaceans	91.8	2135	94.3	9.2			
Amphipoda							
Hyperidae							
<i>Hyperia galba</i>	65.3	309	13.6	6.3	13.5 (2.6)	6.0–20.1	202
<i>Hyperia medusarum</i>	4.1	2	0.1	0.1	17.5 (0.2)	17.3–17.6	2
<i>Hyperoche medusarum</i>	2.0	1	0.0	0.0	7		1
<i>Parathemisto abyssorum</i>	2.0	1	0.0	0.0			
<i>Parathemisto gaudichaudi</i>	2.0	1	0.0	0.0			
Unidentified Parathemisto	4.1	2	0.1	0.0			
Unidentified hyperiids	28.6	19	0.8	0.4			
Gammaridae							
Unidentified gammarids	38.8	56	2.5	0.1	6.7 (1.9)	3.7–9.4	17
Euphausiacea							
Unidentified euphausiids	16.3	14	0.6	1.1			
Decapoda							
Unidentified decapod larvae	6.1	4	0.2	<0.1			
Copepoda	75.5	1724	76.1	1.2			
Unidentified crustaceans	4.1	2	0.1	<0.1			
Cephalopoda	2.0	1	0.0	0.7			
Unidentified squid	2.0	1	0.0	0.7			

Table 4. *Oceanodroma leucorhoa*. Estimates of the diet composition of Leach's storm-petrel using the 3 source (glacier lanternfish, capelin, hyperiid amphipod), dual isotope ($\delta^{15}\text{N}$, $\delta^{13}\text{C}$) linear mixing model described by Phillips & Gregg (2001). Data given as mean \pm SE (95% confidence interval)

	n	% glacier lanternfish	% capelin	% Hyperiid amphipod
Adults				
2001	11	47 \pm 15 (15–78)	5 \pm 7 (0–20)	49 \pm 14 (19–78)
2002	16	87 \pm 15 (56–100)	6 \pm 10 (0–26)	7 \pm 15 (0–38)
Chicks				
2001	10	13 \pm 10 (0–35)	35 \pm 8 (18–52)	51 \pm 11 (28–75)
2002	15	0 \pm 13 (0–29)	21 \pm 10 (1–41)	79 \pm 15 (49–100)

hyperiid amphipod *Hyperia galba* was the most important crustacean in the diet, occurring in 65% of samples and accounting for 6.3% of diet mass. Copepods occurred in 75% of samples but these and euphausiid crustaceans each contributed just only ca. 1% by mass. Unidentified gammarids occurred in 39% of samples, but were also a minor (<0.1%) mass component.

Isotope model estimates of adult and chick diet during breeding

Using isotopic signatures of feathers (corrected), whole fish (*Benthoosema glaciale*, capelin) and amphipods (*Hyperia galba*), we obtained estimates of adult and chick diet from the dual isotope model ($\delta^{15}\text{N}$, $\delta^{13}\text{C}$) of Phillips & Gregg (2001; see Table 4). Toward the end of chick rearing in 2001, we estimate that adult diet consisted of 47% glacier lanternfish, 5% capelin and 49% hyperiid amphipods. Isotope values suggested that in 2002, adults consumed substantially more lanternfish (87%) and less hyperiids (7%), while the contribution of capelin to their diet was similar between years. Adult diet estimated from feather isotope values in 2002 closely resembled that collected via regurgitation that year. Model output for chick diets, however, poorly matched these regurgitations. While 77% of the mass of identified fish in regurgitations were myctophids, feather isotope values suggested that these fish were absent and the chick diet instead consisted of 21% capelin and 79% hyperiid amphipods. Possible reasons for this discrepancy are discussed below.

DISCUSSION

For Leach's storm-petrels breeding at Baccalieu Island in the Northwest Atlantic, we report a typical stepwise trophic enrichment of $\delta^{15}\text{N}$ from amphipods to seabirds. Variation in $\delta^{15}\text{N}$ values among feather

groups suggested both age-related (adult vs. chick) and time of year effects on diet composition and TL, with adults elevating TL during chick rearing and delivering mainly fish to their offspring. Stable carbon isotope values varied among the groups of feathers examined for adult birds, possibly reflecting the latitudinal gradient in $\delta^{13}\text{C}$ (e.g. Rau et al. 1982, Goericke & Fry 1994) as birds shift foraging grounds between breeding and moult (Huntington et al. 1996). Combining conventional and stable isotope approaches has enabled adult–chick TL

comparisons and resulted in some of the first insights into the year-round ecology of Leach's storm-petrel, the smallest and most abundant seabird breeding in eastern North America.

Diet composition and trophic level of Leach's storm-petrels during breeding

As is typical for seabirds, the few dietary studies for Leach's storm-petrel have been conducted during chick rearing when adults return to the colonies with partially digested fish and a mix of crustacean prey for their chicks (Linton 1978, Watanuki 1985, Vermeer & DeVito 1988, Montevecchi et al. 1992). Our results from conventional sampling at Baccalieu late in the 2002 chick-rearing period were consistent with fish, largely myctophids but also capelin and sand lance, comprising the bulk of chick diet, and hyperiid amphipods contributing the remainder. Leach's storm-petrel is highly pelagic during the summer breeding season, feeding up to many hundreds of km from the colony to beyond the edge of the continental shelf (Brown 1986, G. K. Davoren et al. unpubl. data). These distributional patterns are consistent with the myctophid-dominated prey delivered by adults to their chicks. Myctophids or lanternfishes are widespread mesopelagic fish predominantly found in open oceanic waters but also occurring over continental slopes and continental and island shelves (Nafpaktitis et al. 1977). During the day, myctophids occur at 300 to 1200 m, while at night many migrate vertically to feed (Scott & Scott 1988), presumably then becoming available to storm-petrels that feed at the surface. The glacier lanternfish is the most common myctophid north of 40°N in the Northwest Atlantic (Scott & Scott 1988), and it comprised 95% of all myctophids collected in the Newfoundland Basin (McKelvie 1985).

Relative to the offshore distribution and temporally restricted (i.e. nocturnal) availability of myctophid

fishes, *Hyperia galba* is more locally, if seasonally, available to breeding storm-petrels. *H. galba* is a shallow water, parasitic amphipod distributed in Arctic waters in association with large medusae of scyphozoan jellyfish hosts (largely *Aurelia* and *Cyanea*; Dahl 1959, Dittrich 1988). Both *Aurelia* and *Cyanea* occur off the Atlantic coast of Newfoundland (Shih 1977), where *H. galba* is common in fall (Frost 1936). Many amphipods delivered to storm-petrel chicks are intact, sometimes still alive, and are presumably caught close to colonies. In the Gulf of Maine, host jellyfish are common nearshore during summer and fall, seldom being found more than several km from shore (Bigelow 1926). Though highly seasonal (Dittrich 1988), *H. galba* represents a locally available meal for foraging storm-petrels or a 'top-up' for meals of fish collected further afield. Preying on both nearshore crustaceans and offshore fish suggests that Leach's storm-petrels have access to prey conditions that could facilitate a dual short and long foraging trip strategy as found in other procellariids (e.g. Weimerskirch et al. 1994).

Assuming that food loads collected from adults returning to colonies represent meals intended for chicks, there was a surprising lack of correspondence between isotopic and conventional determinations of chick diet in 2002. Model estimates of chick diets were not biologically meaningful because $\delta^{13}\text{C}$ values in their feathers were lower than expected for a diet dominated largely by myctophid fishes. $\delta^{15}\text{N}$ values, however, were within the expected range, and largely similar to those obtained contemporaneously from adult feathers (Table 1). One possible explanation for this discrepancy involves the stomach oil adults feed to chicks, which in 2002 amounted to 43% of chick diet mass. Stable isotope methods trace protein pathways and generally provide little insight into sources of dietary lipids. However, Thompson et al. (2000) proposed that incorporating dietary lipids such as stomach oil (that are relatively depleted in ^{13}C) into protein could provide an explanation for the relatively low $\delta^{13}\text{C}$ values observed in procellariiformes, in general. While this hypothesis remains to be tested experimentally, it provides a potential explanation for our results if storm-petrel adults and chicks differentially rely on stomach oil as an energy source. We recommend measuring stomach oil isotopically in future studies to investigate this possibility.

Focusing on $\delta^{15}\text{N}$ values in feathers, we assessed potential TL separation of adults and chicks and also of adult males and females during breeding, assessments that were simply not possible using conventional sampling techniques. To conduct these comparisons, we assumed similar diet–feather fractionation values for adults and chicks, as a review of data for a variety of animal species (Hodum & Hobson 2000) failed to find

systematic age-related variation. Adult Leach's storm-petrels occupied TL 3.1 in both years (Table 1), equivalent to chicks in 2002 but statistically lower in 2001 when the chick food presumably contained more or higher TL fish. Hodum & Hobson (2000) found significant age-related trophic segregation for 4 Antarctic fulmarine petrel species, with chicks occupying higher trophic positions than adults (TLs were 0.25 to 0.5 higher). Hobson (1993) also found that thick-billed murre *Uria lomvia* and black-legged kittiwake *Rissa tridactyla* chicks in Lancaster Sound, Canada, occupied higher trophic positions than adults, but found no similar separation for northern fulmars *Fulmarus glacialis* or glaucous gulls *Larus hyperboreus*. At least for Leach's storm-petrels in Newfoundland, selectively provisioning chicks with fatty myctophid fishes would facilitate rapid growth, as these fish have higher caloric content than amphipods. Adult–chick dietary differences might also be expected if adults spatially and/or temporally segregate provisioning for themselves and their offspring (Weimerskirch & Cherel 1998, Davoren & Burger 1999, Connan et al. 2005). Our data suggest that adults feed both themselves and their chicks a high proportion of fish during chick rearing, but there may be annual differences in the degree to which parental and offspring diets differ.

We found no significant sex differences in values of either isotope, suggesting that during the chick-rearing period adult males and females feed at a similar TL and use a similar range of foraging locations. For some procellariiform species (e.g. wandering albatrosses *Diomedea exulans* and giant petrels *Macronectes* spp.) substantial sex differences have been reported for both foraging location and the prey species selected for offspring provisioning (Weimerskirch et al. 1993, 1997, Gonzales-Solis et al. 2000, 2002). For these species sex differences have been attributed to the degree of dimorphism which is substantial particularly for the giant petrels. Leach's storm-petrels are largely monomorphic (Huntington et al. 1996), and therefore similar feeding ecologies for males and females are not unexpected.

Feeding ecology over winter: trophic levels and foraging areas

For Leach's storm-petrel, like many other procellariiform seabirds, breeding and moult of the primary feathers are temporally separated to avoid competing needs for energy and nutrients (Ainley et al. 1974, Huntington et al. 1996). Adult TL was significantly elevated (0.25 to 33 higher) during chick-rearing relative to moult when their diet likely consisted of a significant proportion of crustaceans. Adults also fed

upon a wider range of species and occupied a correspondingly wider range of TLs in winter ($\delta^{15}\text{N}$ values ranged 9.9 to 15.7‰, representing TLs 2.1 to 3.7) relative to during the breeding period when their dietary breadth was lower ($\delta^{15}\text{N}$ range 11.3 to 14.8‰, representing TLs 2.4 to 3.5). $\delta^{15}\text{N}$, upon which these comparisons are based, tends to enrich with latitude (Wada et al. 1987), and Leach's storm-petrels winter near the equator (Cramp & Simmons 1977, Huntington et al. 1996). Therefore, while we are fairly confident in the direction of the seasonal dietary change reported here, we may be underestimating its magnitude.

Leach's storm-petrel is highly migratory outside of breeding, moving from colonies in eastern Canada in September/October to winter in the tropics (Cramp & Simmons 1977, Huntington et al. 1996). Migration occurs on both sides of the Atlantic, with some North American birds moving east before going south, evidenced by numbers that far outnumber the European breeding population in the Bay of Biscay region in fall (September to February, with peak numbers in November and December; Hémery & Jouanin 1988). Birds winter off West Africa (Brown 1979). Migration also occurs south along the West Atlantic passing Bermuda in September to November, with birds common offshore of Brazil in winter (Bourne 1959, Cramp & Simmons 1977). In the Southern Ocean lower latitude plankton food bases tend to be enriched in both ^{13}C and ^{15}N relative to higher latitude waters (Wada et al. 1987). The general latitudinal enrichment of $\delta^{13}\text{C}$ (Rau et al. 1982, Goericke & Fry 1994) has been used to elucidate wintering areas of Southern Ocean seabirds from $\delta^{13}\text{C}$ values in their feathers (Cherel et al. 2000, 2002, Quillfeldt et al. 2005). While a latitudinal $\delta^{13}\text{C}$ gradient from the poles to the equator is present in both hemispheres, values in the northern hemisphere encompass a much smaller range (ca. 2‰ in the north vs. 12‰ in the south, Rau et al. 1982). Based upon Rau et al. (1982), the ca. 1‰ $\delta^{13}\text{C}$ enrichment we observed in Leach's storm-petrel primaries during moult suggests a 65° southerly shift, to ca. 10°S. A major assumption inherent in this analysis, however, is that stable isotope ratios in foodwebs follow uniform patterns over vast areas, in this case the entire North Atlantic Ocean, and we can perhaps safely assume this is unlikely. While the observed $\delta^{13}\text{C}$ feather enrichment is likely consistent with a latitudinal effect, insight into the moulting location seems unachievable at present. Future studies as suggested by Hobson (1999) aimed at isotopically characterizing the wintering areas of northern hemisphere migrant seabirds and shorebirds could advance knowledge of trophic relationships and feeding ecology in winter and help to delineate habitat use by seabirds outside the reproductive season.

We observed annual differences in patterns of $\delta^{13}\text{C}$ among primary feathers (Table 1, Fig. 1) that suggests variation in the timing of migration and/or the moult location between years. During the 2000/2001 interbreeding period, there was a clear decline in $\delta^{15}\text{N}$ and TL between early (P1) and mid-moult (P4 and P7), but $\delta^{13}\text{C}$ values were relatively stable, suggesting that all feathers may have been grown at a similar latitude. The 0.5‰ $\delta^{13}\text{C}$ enrichment from mid- to late moult (P10) suggests that birds moved before renewing the final primary. Alternatively, during the 2001/02 moult, $\delta^{13}\text{C}$ values in all primaries were relatively enriched (by 0.7 to 1.0‰) and not dissimilar to those in P10 the previous year. In this year it seems that birds may have renewed all primary feathers closer to the equator.

CONCLUSION

For seabirds, analyzing stable isotope values in feathers grown away from the breeding site is emerging as a powerful method of investigating feeding ecology, trophic relationships and habitat use during winter. Not only are these studies of intrinsic interest for year-round ecology, but providing information about marine bird wintering areas could shed light on both natural (Bertram et al. 2005, Harris et al. 2005) and anthropogenic factors (Wiese et al. 2001, Montevecchi 2002) affecting their survival. Studies aimed at isotopically characterizing wintering areas of northern hemisphere migrant seabird and shorebird species could significantly improve the understanding of winter ecology. Chemical analyses are an efficient way to study the ecology of small species such as Leach's storm-petrels that are unable to carry the currently available archival and telemetric devices.

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