Cytomegalovirus colitis following azacitidine therapy

Rajal Khan MD1, Pam Rudkin PhC2, Kuljit Grewal MD2, Jennifer Leonard MD3, Mowafak Hamodat MD4, Jim Hutchinson MD5, Peter Daley MD5

The present report describes the first recognized case of cytomegalovirus (CMV) colitis following azacitidine therapy. A 66-year-old woman with myelodysplastic syndrome developed CMV colitis, which responded to treatment with ganciclovir. Currently, patients receiving azacitidine do not undergo CMV testing, or receive prophylaxis or CMV-free blood products; however, this policy needs to be revised.

Key Words: Azacitidine; CMV; Colitis

Azacitidine (5-azacytidine, Vidaza [Celgene Corporation, USA]) is a cytidine analogue that was approved in 2004 for the treatment of all myelodysplastic syndrome (MDS) subtypes (1-5). Azacitidine has two main mechanisms of antineoplastic action: cytotoxic activity through the inhibition of nucleic acid synthesis via incorporation into DNA and RNA as an analogue of cytidine, and DNA hypomethylation, and restoration of normal growth control and differentiation of hematopoetic cells (1,2).

The safety of azacitidine has been studied in phase III clinical trials (4). The adverse effects observed were mainly cytopenias; no opportunistic infections were reported (1,2). The drug is not believed to cause significant immunosuppression.

We report a case of cytomegalovirus (CMV) colitis in a patient treated with azacitidine for MDS. To our knowledge, the present report is the first to describe this opportunistic infection in a patient treated with azacitidine.

CASE PRESENTATION

A 66-year-old Caucasian woman was diagnosed with the chronic myelomonocytic leukemia MDS subtype in October 2007, after presenting with pancytopenia, splenomegaly, fatigue and weight loss over a four-month period. Her medical history included spinal stenosis, degenerative disc disease and carpal tunnel syndrome. In August 2009, due to worsening cytopenias and transfusion dependence, the decision was made to begin treatment with azacitidine, starting in September 2009.

The dosing schedule was 75 mg/m² for seven consecutive days, every 28 days. Following monthly doses, her total white blood cell count dropped only slightly, from 2.0 × 10⁹/L to 1.5 × 10⁹/L to 1.7 × 10⁹/L.

On January 21, 2010, after having undergone five cycles of therapy, she presented to the emergency room with fever, abdominal cramping and rectal bleeding. Her urine output was decreased. The patient’s physical examination demonstrated a regular heart rate of 110 beats/min, a blood pressure of 100/50 mmHg and a temperature of 38.4°C. She appeared unwell. Her jugular venous pulsation was weak, and her abdomen was tender in both lower quadrants. There was no hepatosplenomegaly. Her bowel sounds were present, and there was no rebound tenderness. She also experienced mild fecal incontinence. Erythema was present in the perianal area; however, a digital rectal examination was not performed.

Her white blood cell count was 1.0 × 10⁹/L, with an absolute neutrophil count of 0.4 × 10⁹/L and an absolute lymphocyte count of 0.5 × 10⁹/L. Her hemoglobin level was 84 g/L, and platelet count was 71 × 10⁹/L. The patient’s electrolyte, urea, creatinine, lactate dehydrogenase, aspartate aminotransferase, alanine transaminase, alkaline phosphatase, amylase, calcium, albumin and magnesium levels were all within normal limits. Her total bilirubin level was elevated to 29 µmol/L, and her inorganic phosphate level was decreased to 0.70 mmol/L. The patient’s chest radiograph showed mild chronic changes. A computed tomography scan of her abdomen showed “ill-defined thickening of the rectal wall, with minimal stranding within the mesorectal fat and fascia which is associated with increased soft tissue density in the presacral space”.

The patient was admitted to the hematology service with a diagnosis of febrile neutropenia. Blood cultures were drawn, and she was treated with intravenous antibiotics including ceftazidime, metronidazole and imipenem. The patient showed signs of very mild clinical improvement. Her blood cultures were negative after seven days.

Due to continued rectal bleeding, and abdominal and rectal pain, the patient underwent sigmoidoscopy on January 27, which showed the distal 7 cm to 8 cm of her rectal mucosa to be “markedly inflamed and ulcerated” (Figures 1A and 1B). The patient underwent a rectal biopsy, which was reported as “active inflammation with viral effects, compatible with CMV-like colitis”. The CMV immunohistochemical stain was positive (Figure 1C). CMV immunoglobulin G antibody was reactive, CMV immunoglobulin M antibody was nonreactive, CMV shell-vial culture was negative and the CMV viral load was not available at the time of diagnosis. Based on immunohistochemical staining performed on the biopsy tissue, a diagnosis of CMV colitis was made. Intravenous antibiotics were discontinued, and therapy with intravenous ganciclovir was initiated on January 28, 2010, at a dose of 250 mg every 12 h.

At this point, the infectious diseases department was consulted. Azacitidine was held. Repeat sigmoidoscopy was undertaken on February 18, 2010, which showed dramatic improvement in rectal inflammation, with the only abnormality being a small area of...
Chloroquine (seven months ago). Monthly viral loads have been undetectable since the time of discharge, and is no longer dependent on red cell transfusions. To do well on azacitidine treatment, with no further gastrointestinal symptoms, he had a hemoglobin level of 115 g/L and a platelet count of 172 × 10^9/L. This report received no financial support.

erythema in the sigmoid colon (at 25 cm from the rectum) (Figure 1D). Repeat biopsies of the sigmoid colon and rectum showed mild to moderate chronic inflammation with no definite morphological evidence of viral inclusions. Repeat CMV immunohistochemical stains were negative. Ganciclovir was discontinued at this point, after a total duration of three weeks, and the patient was discharged home.

The suggestion of the infectious diseases department was to monitor the serial CMV viral load for viral reactivation if the patient was to be treated further with azacitidine. No CMV suppression was recommended at the time of hospital discharge. The patient has received several more cycles of azacitidine since her discharge from the hospital. Most recent bloodwork showed a white blood cell count of 3.6 × 10^9/L, with an absolute neutrophil count of 1.5 × 10^9/L, hemoglobin level of 115 g/L and a platelet count of 172 × 10^9/L. She continues to do well on azacitidine treatment, with no further gastrointestinal symptoms, and is no longer dependent on red cell transfusions. Monthly viral loads have been undetectable since the time of discharge (seven months ago).

**DISCUSSION**

We describe a case of CMV colitis in a 66-year-old woman treated with azacitidine for MDS. To our knowledge, this is the first report of this opportunistic infection in a patient treated with this medication. Adverse events have been reported with the use of azacitidine including nonspecific gastrointestinal symptoms (nausea, vomiting, diarrhea, constipation and anorexia), hematological events (neutropenia, anemia and thrombocytopenia), injection site events, arthralgia, cough, dyspnea, headache, dizziness, weakness and insomnia. Reported hospitalizations were from thrombocytopenia, febrile neutropenia, fever and pneumonia. However, none of these symptoms have been attributed to CMV infection (1,2).

In our patient, there was a dramatic response, both clinically and endoscopically, to treatment with intravenous ganciclovir. No formal guidelines for duration of therapy exist in this circumstance. Our decision to treat for three weeks was based on the patient’s clinical and endoscopic response to ganciclovir treatment.

It is not clear, in the present case, whether there was latent CMV infection, which was reactivated by azacitidine, or whether primary infection with CMV occurred after initiation of azacitidine treatment because CMV serology was not performed before azacitidine treatment. Azacitidine is known to activate CMV transcription promoters in vitro (6) and in vivo (7). In fact, the drug is used to facilitate gene transfer of viral vectors for cancer treatment (8), which is believed to occur through the drug’s effect on DNA promoter demethylation and activation of the nuclear factor-kappa B pathway. Whether azacitidine used for treatment of MDS has any effect on reactivation of latent CMV in MDS patients is unknown.

Although severe CMV infections, including colitis (9), can affect immunocompetent hosts (10), it is generally an infection that occurs during profound immunosuppression including late-stage HIV infection (11) and following bone marrow transplant (12). Generally, conventional chemotherapies that do not cause lymphopenia or alterations in lymphocyte function are not considered to be high risk for the development of CMV disease (13,14). However, isolated cases of CMV colitis have been reported after conventional chemotherapy for testicular cancer (15), non-Hodgkin’s lymphoma (16), hypopharyngeal cancer (17), pancreatic carcinoma (18) and small cell lung cancer (19). CMV disease of other organ systems after conventional chemotherapy has also been reported (20). Furthermore, a recent study (21) suggested that there may be a high incidence of CMV reactivation in patients undergoing conventional chemotherapy.

The optimal approach to prevention in compromised hosts is not clear because both prophylactic and pre-emptive treatment approaches have advantages (22). There is no existent policy in Canada for CMV screening before initiation of azacitidine therapy, or for CMV prophylaxis in patients known to have chronic CMV infection while being treated with azacitidine. Patients on azacitidine are not generally given CMV-free transfusions. Further study of the relationships among azacitidine, CMV promoter activation, and acute or reactivated CMV infection are needed to answer these questions. We suggest that CMV testing, monitoring and prophylaxis may be required in patients receiving azacitidine.

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**REFERENCES**


